

Risperidone attenuates brain damage after focal cerebral ischemia in vivo

Burak Yulug^{a,*}, Aysegül Yildiz^b, Orkide Güzel^c, Erdinc Kilic^e,
Wolf Rüdiger Schäbitz^d, Ertugrul Kilic^e

^a Department of Neurology, University of Uludag, Bursa, Turkey

^b Department of Psychiatry, University of Dokuz Eylül, 35340 Izmir, Turkey

^c Department of Pediatric Neurology, University of Dokuz Eylül, 35340 Izmir, Turkey

^d Department of Neurology, University, Hospital Münster, 48149 Münster, Germany

^e Department of Neurology, University Hospital, Zurich, 8091 Zurich, Switzerland

Received 27 December 2005; received in revised form 11 March 2006; accepted 20 March 2006

Available online 18 April 2006

Abstract

Since their introduction, atypical neuroleptic agents have been discovered to have some beneficial effects beyond their effectiveness as neuroleptic drugs. Among these initially unexpected effects are their potential effects as mood stabilizers in bipolar disorder and their efficacy in improving long-term outcome in schizophrenia. These effects recently raised the question whether these drugs may also have some neuroprotective effect in the brain. To examine this matter, in this study we evaluated the neuroprotective effect of risperidone after permanent focal cerebral ischemia. Anaesthetized male C57BL/6j mice were submitted to permanent thread occlusion of the middle cerebral artery (MCA). Risperidone (0.1, 1 or 10 mg/kg) or vehicle was applied intraperitoneally just after permanent ischemia.

Twenty-four hours after permanent ischemia, brain injury was evaluated by triphenyltetrazolium chloride staining (TTC). Risperidone (0.1, 1 and 10 mg/kg) showed significant neuroprotection after permanent focal cerebral ischemia.

© 2006 Elsevier Inc. All rights reserved.

Keywords: Risperidone; Cerebral ischemia; Neuroprotection; Stroke

1. Introduction

Stroke is the third leading cause of death and adult morbidity in developed countries [1,2]. Many deleterious cellular pathways have been proposed to explain the molecular pathogenesis of this clinically devastating disease [3]. Interest in role of neurotransmitters in the pathogenesis of ischemic stroke resulted in studies establishing that release of not only glutamate but also other neurotransmitters (i.e. dopamine and serotonin) and their excitotoxic actions are important for the development of ischemic neuronal damage [4,5]. In this respect, the extracellular release of dopamine and serotonin is shown to be correlated with the extent of the ischemic damage [6,7].

Despite serious progresses in basic neuroscience research, which focused especially on the in vitro development of new neuroprotective compounds, the translation of these research findings into therapeutic advances for neurological diseases is frustratingly slow. Thus, clinically applicable and experimentally neuroprotective drugs should be studied more than before, i.e. erythropoietin in clinical stroke [8].

Risperidone is one of the member of the atypical neuroleptic drugs and a candidate as a clinically applicable neuroprotective drug. In addition to its well-known antidopaminergic and antiserotonergic effects, risperidone is also shown to stimulate a two- to three-fold increase in newly divided cells in the rat subventricular zone [9].

It is well known that the atypical neuroleptic drugs are widely used in the treatment of schizophrenia and bipolar disorder. These agents, in addition to their superior efficacy profile relative to conventional neuroleptic agents for acute treatment of psychotic symptoms are also considered to be more effective

* Corresponding author. Tel.: +90 535 7023338; fax: +90 232 4468090.
E-mail address: yulug@gmx.de (B. Yulug).

for improving long-term outcome in schizophrenia [10–14]. Schizophrenia is also now considered to be a neurodevelopmental and neurodegenerative disease according to the clinical and neuroimaging studies [14–17]. These results indicate that atypical neuroleptics may have a mechanism of action other than the previously proposed mechanisms, which might explain their role in improved cognition in animals and in schizophrenic patients. Altogether these findings suggest that atypical antipsychotics with their potential to stimulate neuronal replacement and repair may also be promising in other neurodegenerative diseases.

On the basis of insufficient safety and limited therapeutic time window of thrombolytic therapy in humans, we thought that the permanent focal cerebral ischemia model would have more clinical relevance than the transient cerebral ischemia in this study. However, in addition to the well-known antidopaminergic and antiserotonergic effects of risperidone, the evidences about its neuroprotective effects are rapidly replicating. These factors prompt us to choose risperidone as a pharmacological agent, whereas, it was expected that its antidopaminergic and antiserotonergic effects would be enough to prevent or delay ischemia in a relevant model for the nonthrombolized stroke in humans.

In our study, we have examined the effect of risperidone in the mouse model of focal permanent cerebral ischemia and administered risperidone just after production of permanent ischemia. Reproducibility of cerebral ischemia was controlled by Laser Doppler Flowmetry and the neuroprotection was evaluated by infarct volume measurements.

2. Materials and methods

2.1. Experimental groups

All experimental procedures were carried out with governmental approval according to local guidelines for the care and use of laboratory animals. Adult male C57BL/6j mice weighing 21–25 g was assigned to the following experiments and groups: Intraperitoneal administration of (a) 0.1 mg/kg risperidone, (b) 1 mg/kg risperidone, (c) 10 mg/kg risperidone or (d) 0.2 ml vehicle (0.09% NaCl), starting at the onset of 24 h of permanent focal cerebral ischemia ($n = 6$ animals/group). Risperidone (Janssen Cilag Pharmaceuticals) were dissolved in 0.1 N HCl and then buffered with NaOH (final pH 7.1). By referring to the recommended clinical dose range of 4–6 mg/day of risperidone in humans and 70–100 times higher metabolic rate in mice we determined averaged doses of risperidone as 0.1, 1 and 10 mg/kg [18].

2.2. Induction of ischemia

Animals were anesthetized with 1% halothane (30% O₂, remainder N₂O). Rectal temperature was maintained between 36.5 and 37.0 °C using a feedback-controlled heating system. During the experiments, cerebral blood flow was measured by laser Doppler flowmetry (LDF) using a flexible 0.5 mm fiber optic probe (Perimed, Stockholm, Sweden), which was attached to the intact skull overlying the MCA territory (2 mm posterior/6 mm lateral from bregma). LDF changes were monitored up to 30 min after the onset of ischemia. Focal cerebral ischemia was induced using an intraluminal filament technique [19,20]. A midline neck incision was made, and the left common and external carotid arteries were isolated and ligated. A microvascular clip (FE691, Aesculap, Tuttingen, Germany) was temporarily placed on the internal carotid artery. A 8-0 nylon monofilament (Ethilon; Ethicon, Norderstedt, Germany) coated with silicon resin (Xantopren, Bayer Dental, Osaka, Japan; diameter of the coated thread: 190–200 μm) was introduced through a small incision into the common carotid artery and advanced 9 mm distal to the carotid bifurcation for permanent occlusion of the MCA. Anesthesia was discontinued and animals were placed into their home cages.

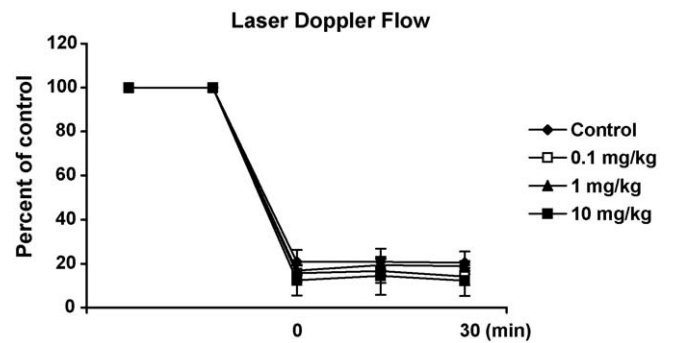


Fig. 1. Laser Doppler flow measurements (LDF) above middle cerebral artery territory during permanent intraluminal thread occlusion in animals treated with vehicle solution or risperidone. No differences in LDF values were detected between animal groups. Values are means \pm S.D.

tion of the MCA. Anesthesia was discontinued and animals were placed into their home cages.

2.3. Triphenyltetrazolium chloride (TTC) staining

Animals were reanesthetized with overdose halothane and decapitated. Brains were incubated for 5 min in ice-cold isotonic saline and coronally cut into five 2 mm slices using a mouse brain matrix (BRM-2000C; Activational System Inc., MI, USA). These slices were immediately stained with 2% 2,3,5-triphenyltetrazolium chloride (TTC) for 20 min. The border between infarcted and non-infarcted tissue was outlined using an image analysis system, and the area of infarction was measured by subtracting the area of the non-lesioned ipsilateral hemisphere from that of the contralateral side. The infarct volume was calculated by integration of the lesion areas [19–21].

2.4. Statistics

All values are given as mean \pm S.D. Differences between groups were compared by using oneway ANOVA analysis followed by LSD tests after permanent cerebral ischemia. P -values < 0.05 were considered to indicate statistical significance.

3. Results

3.1. Laser Doppler Flow (LDF)

Mean LDF reproducibly declined to $< 15\%$ of pre-ischemic control levels immediately after thread insertion in all animal groups. No differences were seen between various animal groups (Fig. 1).

3.2. Triphenyltetrazolium chloride (TTC) staining: infarct size

Reproducible brain infarcts were obtained in the 0.1, 1 and 10 mg/kg of risperidone or 0.2 ml of vehicle treated groups (Fig. 2). Intraperitoneal administration of 0.1, 1 and 10 mg/kg risperidone significantly reduced the infarct volume as compared with control animals (Fig. 2).

4. Discussion

In the ischemic brain it is well known that a massive release of dopamine can amplify the neuronal damage caused by exci-

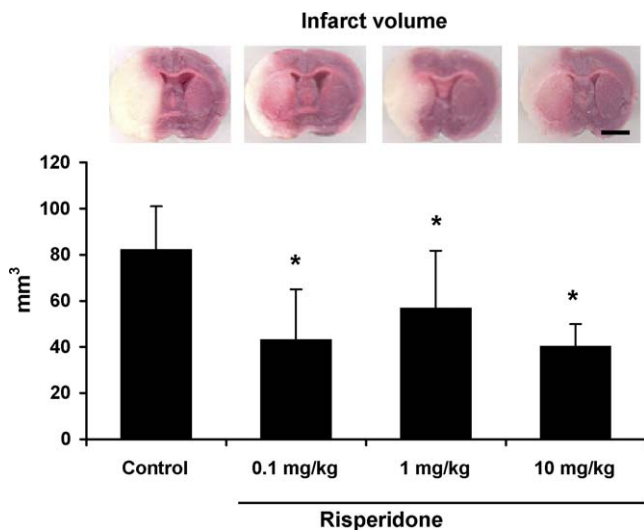


Fig. 2. Infarct volumes of mice subjected to permanent cerebral ischemia. Animals treated with risperidone (0.1, 1 or 10 mg/kg). Note that 0.1, 1 and 10 mg/kg risperidone lead to a decrease in infarct size. *Significantly different from vehicle-treated control animals ($P \leq 0.05$). Data are given as means \pm S.D.

toxicity and energy deprivation [22–24]. Besides dopamine, serotonin is also shown to modulate the postsynaptic effects of glutamate and leads to reduction of blood flow during cerebral ischemia [25,26]. Therefore, ischemia-induced abnormal release of such neurotransmitters in vulnerable brain areas may represent a critical factor that transforms a transient ischemic attack into an ischemic episode resulting in irreversible consequences in brain tissue and its synaptic circuits.

In this respect risperidone via its antidopaminergic and anti-serotonergic effects would be expected to prevent or improve ischemia. Given also the neuroprotective effects of the classical mood stabilizers “lithium and valproate” [27–29], clinical evidence suggesting some mood stabilizing effects of the risperidone would provide further rationale for investigating possible neuroprotective effects of this agent. Indeed, other studies have documented that atypical antipsychotics such as quetiapine, olanzapine and clozapine modulate brain derived nerve growth factor (BDNF) expression and B cell lymphoma protein (bcl-2) levels in mouse and rat brain after chronic exposure [30–32].

In contrast to a recent study [33], in which pretreatment with risperidone 7 days before focal cortical photothrombosis did not affect histological outcome in aged rats, we observed that acute administration of 0.1, 1 and 10 mg/kg of risperidone significantly decreased the infarct size in permanent focal cerebral ischemia. In higher dose group we have not observed an additional decrease of infarct size in the risperidone 10 mg/kg dose group. It is difficult to estimate what caused that difference without getting the results from further experiments on the anti-apoptotic effects of various atypical antipsychotic agents with different receptor affinities. However, there is data indicating enhancing effect of adrenergic activity in neuroprotection as well as reversal of such effect under adrenergic blockage [34–37]. This together with the knowledge that risperidone at higher doses exerts anti-adrenergic activity may explain the failure of 10 mg/kg of risperidone to show an additional neuroprotection.

To the extent that these data in mice translate to people, it is unlikely that risperidone worsens neuronal damage caused by small strokes, which elderly patients probably sustain frequently. This finding is interesting in light of recent clinical evidence of an increased rate of cerebrovascular events in risperidone-treated dementia patients [38–40]. It was found that 4% of risperidone treated dementia patients suffered cerebrovascular adverse events compared with 2% in placebo-treated patients [40]. Because risperidone provided significant neuroprotection in mice, it might trigger other mechanism(s), including vascular pathology, known to be associated with higher stroke risk in demented patients [41].

Additionally, it is widely known that occurrence of post-stroke mood disorders, especially depression, is one of the most frequent complications of stroke [42]. It affects approximately 20–40% of all patients and the definitive treatment of this clinical picture includes various antidepressant agents [43]. Moreover, besides its better tolerability than the typical antipsychotic agents and additional mood stabilizing effect, its actually proven neuroprotective property by our study could also provide strong preclinical evidence to its role as a possible candidate for post-stroke mood disorders.

In summary, this study provides evidence that acute administration of risperidone by the dose range indicated protects neurons from death after permanent cerebral ischemia. This finding indicates that atypical neuroleptic agents in common with lithium and valproate may have neuroprotective actions. Further experiments to evaluate the long-term clinical reflections of such neuroprotective effects of atypical neuroleptics also in bipolar/schizophrenic patients via magnetic resonance imaging and spectroscopy studies would be the logical future steps to be taken in the field of psychiatric research.

References

- [1] C.L.P. Sudlow, C.P. Warlow, International Stroke Incidence Studies Collaboration, Comparable studies of the incidence of stroke and its pathological types: results from an international collaboration, *Stroke* 28 (1997) 491–499.
- [2] P.M. Rothwell, The high cost of nonfunding stroke research: a comparison with heart disease and cancer, *Lancet* 357 (2001) 1612–1616.
- [3] U. Dirnagl, C. Iadecola, M.A. Moskowitz, Pathobiology of ischaemic stroke, An integrated view, *Trends Neuroscience* 22 (1999) 391–397.
- [4] B. Meldrum, Excitotoxicity in ischemia, An overview, in MD Gingsberg, WD Dietrich (Eds.), *Cerebrovascular Diseases: Sixteenth Research (Princeton) Conference*, Raven Press Publishers, New York, 1989, pp. 47–60.
- [5] M.Y.T. Globus, R. Busto, W.D. Dietrich, E. Martinez, I. Valdes, M.D. Gingsberg, Effect of ischemia on in vivo release of striatal dopamine, glutamate and gamma aminobutyric acid studies by intracerebral microdialysis, *J. Neurochem.* 51 (1988) 1455–1464.
- [6] D.A. Richards, T.P. Orenovich, L. Symon, G. Curzon, Extracellular dopamine and serotonin in the rat striatum during transient ischemia of different severities: a microdialysis study, *J. Neurochem.* 60 (1993) 18–136.
- [7] N. Hashimoto, T. Matsumoto, H. Mabe, T. Hashitani, H. Nishino, Dopamine has inhibitory and accelerating effects on ischemia induced neuronal cell damage in the striatum, *Brain Res. Bull.* 33 (1994) 281–288.
- [8] H. Ehrenreich, M. Hasselblat, C. Dembowski, L. Cepek, P. Lewczuk, M. Stiefel, H.H. Rustenbeck, N. Breiter, S. Jakob, F. Knerlich, M. Bohn,

- W. Poser, E. Ruther, M. Kochen, O. Gefeller, C. Gleiter, T.C. Wessel, M. De Ryck, L. Itri, H. Pange, A. Cerami, M. Brines, A.L. Siren, Erythropoietin therapy for acute stroke is both safe and beneficial, *Mol. Med.* 8 (2002) 495–505.
- [9] C.G. Wakade, S.P. Mahadik, J.L. Waller, F.C. Chiu, Atypical neuroleptics stimulate neurogenesis in adult rat brain, *J. Neurosci. Res.* 69 (2002) 72–79.
- [10] J.L. Waddington, E. O'Callaghan, P. Buckley, C. Larkin, O. Redmond, J. Stack, J.T. Ennis, The age differences of MRI abnormalities in schizophrenia suggests early ventricular enlargement but later prominence of cortical atrophy, *Schiz. Res.* 5 (1991) 188–189.
- [11] R.J. Wyatt, Early intervention with neuroleptics may decrease the long term morbidity of schizophrenia, *Schiz. Res.* 5 (1991) 201–202.
- [12] R.J. Wyatt, I.D. Henter, The effects of early and sustained intervention on the long-term morbidity of schizophrenia, *J. Psychiatr. Res.* 32 (1998) 169–177.
- [13] I.R.H. Falloon, Early intervention for first episodes of schizophrenia, A preliminary exploration, *Psychiatry* 55 (1992) 4–15.
- [14] Le. DeLisi, M. Sakuma, W. Rew, M. Kushner, A.L. Hoff, R. Grimson, Schizophrenia as a chronic active brain process: a study of progressive brain structural change subsequent to the onset of schizophrenia, *Psychiatry Res. (Neuroimaging Section)* 74 (1997) 129–140.
- [15] E.C. Johnstone, C.D. Frith, J. Husband, L. Kreef, Cerebral ventricular size and cognitive impairment in chronic schizophrenia, *Lancet* 11 (1996) 924–926.
- [16] D.L. Garver, The etiologic heterogeneity of schizophrenia: review and perspective, *Harvard Rev. Psychiatr.* 4 (1997) 1–11.
- [17] J.A. Lieberman, Pathophysiologic mechanisms in the pathogenesis and clinical course of schizophrenia, *J. Clin. Psychiatr.* 60 (1999) 9–12.
- [18] J.R. Speakman, D.A. Talbot, C. Selman, S. Snart, J.S. McLaren, P. Redman, E. Krol, D.M. Jackson, M.S. Johnson, M.D.I. Brand, Uncoupled and surviving: individual mice with high metabolism have greater mitochondrial uncoupling and live longer, *Aging Cell* 3 (2004) 87–95.
- [19] B. Yulug, U. Kilic, E. Kilic, M. Bahr, Rifampicin attenuates brain damage in focal ischemia, *Brain Res.* 996 (2004) 76–80.
- [20] E. Kilic, G.P.H. Dietz, D.M. Hermann, M. Bähr, Intravenous delivery of TAT-Bcl-XL fusion protein is protective before and after middle cerebral artery thread occlusion in mice, *Ann. Neurol.* 52 (2002) 617–622.
- [21] E. Kilic, J. Weishaupt, U. Kilic, G. Rohde, B. Yulug, K. Peters, D.M. Hermann, M. Bähr, The superoxide dismutase1 (sod1) g93a mutation does not promote neuronal injury after transient focal brain ischemia and optic nerve transection in mice, *Neuroscience* 128 (2004) 359–364.
- [22] T. Brannan, J. Weinberger, P. Knott, I. Taff, H. Kaufmann, D. Togasaki, J. Nieves-Rosa, H.L. Maker, Direct evidence of acute, massive striatal dopamine release in gerbils with unilateral strokes, *Stroke* 18 (1987) 108–110.
- [23] D.J. Surmeier, J. Bargas, H.C. Hemmings, A.C. Nairn, P. Greengard, Modulation of calcium currents by D1 dopaminergic protein kinase/phosphatase cascade in rat neostriatal neurons, *Neuron* 14 (1995) 385–397.
- [24] F.H. Khan, M. Saha, S. Chakrabarti, Dopamine induced protein damage in mitochondrial-synaptosomal fraction of the brain, *Brain Res.* 895 (2001) 245–249.
- [25] J.F. Reinhard, J.E. Liebmann, A.J. Schlosberg, M.A. Moskowitz, Serotonin neurons project to small blood vessels in the brain, *Science* 206 (1979) 85–87.
- [26] S. Nedegard, I. Engberg, J.A. Flatman, The modulation of excitatory amino acid responses by serotonin in the cat neocortex in vitro, *Cell Mol. Neurobiol.* 7 (1987) 367–379.
- [27] F.J. Vajda, Valproate and neuroprotection, *J. Clin. Neurosci.* 9 (2002) 508–514.
- [28] M.R. Jeong, R. Hashimoto, V.V. Senatorov, K. Fujimaki, M. Ren, M.S. Lee, D.M. Chuang, Valproic acid, a mood stabilizer and anticonvulsant, protects rat cerebral cortical neurons from spontaneous cell death: a role of histone deacetylase inhibition, *FEBS Lett.* 542 (2003) 74–78.
- [29] D.M. Chuang, Neuroprotective and neurotrophic actions of the mood stabilizer lithium: can it be used to treat neurodegenerative diseases? *Crit. Rev. Neurobiol.* 16 (2004) 83–90.
- [30] O. Bai, J. Chlan-Fourney, R. Bowen, D. Keegan, X.M. Li, Expression of brain-derived neurotrophic factor mRNA in rat hippocampus after treatment with antipsychotic drugs, *J. Neurosci. Res.* 71 (2003) 127–131.
- [31] O. Bai, H. Zhang, X.M. Li, Antipsychotic drugs clozapine and olanzapine upregulate bcl-2 mRNA and protein in rat frontal cortex and hippocampus, *Brain Res.* 1010 (2004) 81–86.
- [32] Z. Wei, D.D. Mousseau, J.S. Richardson, L.E. Dyck, X.M. Li, Atypical antipsychotics attenuate neurotoxicity of beta-amyloid(25–35) by modulating Bax and Bcl-X(l/s) expression and localization, *J. Neurosci. Res.* 74 (2003) 942–947.
- [33] C.S. Zhao, K. Puurunen, T. Schallert, J. Sivenius, J. Jolkkonen, Behavioral and histological effects of chronic antipsychotic and antidepressant drug treatment in aged rats with focal ischemic brain injury, *Behav. Brain Res.* 30 (2005) 211–220.
- [34] W.E. Hoffman, M.A. Cheng, C. Thomas, V.L. Baughman, R.F. Albrecht, Clonidine decrease plasma catecholamines and improves outcome from incomplete ischemia in the rat, *Anesth. Analg.* 73 (1991) 460–464.
- [35] W.E. Hoffman, E. Kochs, C. Werner, C. Thomas, R.F. Albrecht, Dexmedetomidine improves neurologic outcome from incomplete ischemia in the rat: Reversal by the α_2 -adrenergic antagonist atipamezole, *Anesthesiology* 75 (1991) 328–332.
- [36] C. Maier, G.K. Steinberg, G.H. Sun, G.T. Zhi, M. Maze, Neuroprotection by the alpha sub 2-adrenoceptor agonist dexmedetomidine in a focal model of cerebral ischemia, *Anesthesiology* 79 (1993) 306–312.
- [37] Y. Zhang, Clonidine preconditioning decreases infarct size and improves neurological outcome from transient forebrain ischemia in the rat, *Neuroscience* 125 (2004) 625–631.
- [38] W.V. Bobo, K.F. More, Potential association between risperidone and cerebrovascular events. *Prim Care Companion, J. Clin. Psychiatry* 5 (2003) 141.
- [39] D.A. Smith, M.T. Beier, Association between risperidone treatment and cerebrovascular adverse events: examining the evidence and postulating hypotheses for an underlying mechanism, *J. Am. Med. Dir. Assoc.* 5 (2004) 129–132.
- [40] E. Wooltorton, Risperidone (Risperdal): increased rate of cerebrovascular events in dementia trials, *CMAJ* 167 (2002) 1269–1270.
- [41] J. Shi, G. Perry, M.A. Smith, R.P. Friedland, Vascular abnormalities: the insidious pathogenesis of Alzheimer's disease, *Neurobiol. Aging* 21 (2000) 357–361.
- [42] S. Paolucci, G. Antonucci, M.G. Grasso, D. Morelli, E. Troisi, P. Coiro, Post-stroke depression, antidepressant treatment and rehabilitation results, A case-control study, *Cerebrovasc. Dis.* 12 (2001) 264–271.
- [43] E.M. Whyte, B.H. Mulsant, Post stroke depression: epidemiology, pathophysiology and biological treatment, *Biol. Psychiatry* 52 (2002) 253–264.